KEY TO FREQUENTLY NAMED LEPIDOPTERAN LARVAE INTERCEPTED, OR POTENTIALLY ENCOUNTERED, AT US PORTS

S. C. Passoa, 2014

This key is designed to identify the most frequently named Lepidoptera at United States ports of entry as of 2013. Because many species cannot be named (early instars or poorly studied groups), it is not a given that this key has all the most frequently intercepted taxa. Some genera may regularly intercepted but unrecognized. The huge variety of early instar Noctuidae/Erebidae is a good example of this problem. There are many other taxa in this category. Trade patterns are constantly changing over time, users should expect a need to delete or add species to this key in the future.

Good comprehensive larval keys exist (Carter and Kristensen 1998, Stehr 1987) but as a rule they are too long and complicated for the volume of material we get in APHIS. Also, they rarely go past family. This key is a compromise between the need to for precision and speed. Thus, a good collection and detailed understanding of larval morphology is assumed and required. All of the references cited in the LepIntercept fact sheets need to be part of any port library. This document serves as a blanket recommendation for their purchase or copying costs from the APHIS Lepidoptera specialist to any port needing a justification. This key assumes eventual full access to the appropriate literature. Getting this information needs to be a priority for ports frequently intercepting larval Lepidoptera. Many of the copyrighted books are never going to be "on-line" and go out of print relatively fast. Aggressive hassle free purchasing is a must for any serious identifier having to deal with Lepidoptera.

Weisman (1974, 1986) and the Pest ID System – previously called PIN, PINET) database were consulted to select species for this key. Additions were made based on the experience of the author. For every species in LepIntercept, we include a diagnosis and reference the appropriate data sheet. To increase taxon coverage, several other categories have been added. Some species are commonly seen at the ports but were outside the scope of our study (not in the top 80%). They are mentioned to point identifiers in the right direction. When there is a relatively easy diagnosis, it is included. A third category is composed of major pests not commonly intercepted, but serious enough to warrant inclusion in the key. This usually involved consulting the Pests Not Known to Occur in the United States (PNKTO) series and the Off Shore Pest Information System list for Lepidoptera (http://www.aphis.usda.gov/plant_health/safeguarding/ downloads/2011/OffshorePestInformationSystemPestListProject.pdf). Studies on quarantine Lepidoptera have produced new species and novel morphological characters. United States ports do not just intercept "common species". Some rare larvae, if they would cause confusion, are added. Very distinctive larvae.

"common species". Some rare larvae, if they would cause confusion, are added. Very distinctive larvae are also sometimes included. With a few exceptions, if a genus or species is mentioned, identifiers can expect these taxa can be recognized with morphology. Pupae are rarely intercepted but are included in this key if they have been found in significant numbers over several years.

Unlike traditional taxonomic keys to a given category (genus, species, etc.), this key mixes family to

Unlike traditional taxonomic keys to a given category (genus, species, etc.), this key mixes family to species identifications and adds "possibly" or "usually" or "probably" if needed. These warnings emphasize the importance of confirming the identification with additional literature, morphology and host/origin data. Many of the taxa categorized as "probably" or "usually" could be confirmed with more effort but their diagnosis was too complicated to include as a bullet statement. They appear in the index with a question mark. This key suggests other possibilities where better identifications are often possible for areas with more complete literature. Thus, the key stops at Sphingidae, but one should not assume that no larval sphingid can ever be identified. Rather, there was no common pattern of species that could be suggested.

Do not forward specimens to specialists with no data (origin unknown and no host) because these contribute nothing to the Pest ID database unless the pest is especially significant (e.g., a high ranking OPIS species). Those missing parts (head or abdomen) should also be held because the chance of getting a good identification on such material is slim.

The following works (and references within them) have been consulted freely to construct this key: Weisman (1974, 1986), Epstein (1996), Stehr (1987), Passoa (2007), Kristensen (1998) and Schnitzler et al. (2012). The pyraloid section was based on Weisman (1974, 1986, 1987), Passoa (1985, 2007), Solis (2011 revision) and Hayden et al. (2013). The Noctuoidea was modified from Weisman (1986), Crumb (1956), Miller (1991), and Passoa (2007). The Tortricidae key was authored by Todd Gilligan. Consult Gilligan and Epstein (2012) for interactive identification keys covering additional tortricid species of quarantine significance. The Gelechiidae species list and index is by Jim Young. All the references in the fact sheets and numerous unpublished APHIS training aids were also consulted.

No attempts were made to place the taxa in phylogenetic order nor were characters studied to decide the best way to organize the key. Families were eliminated one by one based on how easy the taxon was to define or how common the species are at the ports. The organization generally follows Weisman (1986) and aims for an almost equal number of couplets. Better versions and schemes with illustrations are already being considered and tested for the future. Classification of the Lepidoptera follows Nieukerken et al. (2011).

Characters ordered from head to anal shield without regard to importance. Terms are defined in Stehr (1987). The most obvious terminology change from Weisman (1986) is that stemmata is used in lieu of ocelli. This key does not have any "escape couplets" (for example, none of the above) except in Noctuidae/Erebidae. Therefore it is important not to force an answer. If nothing fits, it is likely the identifier has a species not covered by this key. The characters used to identify the included taxa are often suitable for situations outside the ports. If *Spodoptera frugiperda* is suspected in corn from North America, the key and fact sheets can help confirm or refute the identification. The reverse is not true. Keys for intercepted Lepidoptera are not designed to identify random unknown larvae outside the ports. This situation was pointed out by Solis (2011) and it still remains a valid warning. Be careful when only a host and scientific name is given without a diagnosis. This is a good place to start but there usually are other possibilities. Where it has been studied (pyraloids and Noctuidae), it is painfully obvious that early instar larvae often will not key to species correctly. Thus, most early instar larvae should not be forwarded to specialists because morphological identification is not possible at this time.

If there are questions, do not just pop the larva in the mail and expect a specialist to read long notes or guess at the issue. Instead, take a photograph of the structure and email it. This allows for mutual viewing of the segment in question and almost always will lead to a fast clarification. Some Lepidoptera are urticating, port identifiers need to point this out to less experienced inspectors. The caterpillar fauna of Japan, Australia, Europe, Canada and North America are best known. Africa is the least studied. Asia and Latin America are partially documented. This should be kept in mind when deciding to forward a specimen to a specialist. No effort was made to repeat large sections of keys that required no changes. Pierids on crucifers, tineids and phycitines on stored products, and New World *Spodoptera* are some examples.

This key points out significant problems in identification of quarantine Lepidoptera and summarizes the "state of the art." Nearly all these problems could be overcome with time to carefully study specimens using a large series of known larvae in all the instars. Worldwide, quarantine workers feel pressured to get enough time outside their identification load and collections almost always lack the needed material. It is wrong to conclude "larvae can't be identified"; results come after resources are allocated. Systematics is no different from any other scientific endeavor in this regard.

There are many caveats to this key. Most serious, few interception records are based on barcoded larvae or are associated with reared adults. Their hosts are never vouchered and many plant identifications are not made by trained botanists. Origins can be misleading because of shipping changes in transit. All of these factors must be remembered when analyzing Lepidoptera intercepted at ports of entry. The reliability of any association is based on repetition. The more times a species is found on a given host from a given pathway, the more likely that association is real and valid.

Given a very short time frame, a large series of specimens could not be examined. There was no time to carefully cite literature for all the facts presented. No effort was made to separate literature records from recent or past interception data for species not covered in the fact sheets. The Pest ID database provides the most up to date listing of hosts and orgins. No effort was made to duplicate this data with additional indices or lists although interception data was critically evaluated for species covered in the fact sheets. Nevertheless, it is expected that this key will produce a correct identification for a large majority of Lepidoptera intercepted at United States ports when used in combination with the fact sheets. The Systematic Entomology Laboratory deserves credit for producing the taxonomic monographs fundamental to any key. APHIS port identifiers answered questions, took pictures and donated specimens. Hopefully this key repays their efforts and patience. Any further comments or corrections are welcome.

1. Body with numerous secondary setae or scoli, hiding most of the primary setae2

Secondary setae can be spinelike (e.g. Arctiinae) or in arranged in tufts (e.g. Lymantriinae). Occasionally the secondary setae are minute and the larva appears smooth even though the body is covered with hairs (e.g. Papilionidae). Scoli are sometimes present. Some Arctiinae erebids, such as *Utethesia* on *Crotolaria*, are an exception. They best fit here because the bisetose L 3 group on A1-8 can be conisidered secondary setae in a regular pattern, even though the covering is sparse. Minute spinules on the skin do not count as secondary setae (Heliothinae, etc.).

1'. Body without numerous short secondary setae or scoli, primary setae evident12

Some gelechioids have secondary setae on the prothoracic shield (Peleopodidae). A genus of sod webworms has scattered secondary setae on the anal prolegs. *Anarsia* has secondary setae on the anal shield. Also, a few families have one or two "extra setae" (Cossidae, Thyatiridae, some Tortricidae, seta X of the Notodontidae and a few Noctuidae). All these examples should be ignored for this couplet.

2. Head usually much larger than prothorax, body widest at middle; lenticles present; crochets biordinal or triordinal in a laterally elongated circle........HESPERIIDAE

Two of the three hesperiid subfamilies have a "neck". All subfamilies have lenticles. Minno (1994) has a larval key to some New World pest species. APHIS is most likely to see *U. proteus*. From at least 1989 or earlier, the maguey worm *Aegiale hesperiaris* (or more rarely, *Agathymus* sp.) is no longer the most common worm in bottles of tequila. Instead these are being replaced with the cossid *Comadia* (see couplet 49, Brickey and Gorharn 1989) Some hesperiid species have an anal fork. The abdominal segments are indistinctly annulated. Distribution: worldwide.

and hidden; body sometimes sluglike; lenticles usually absent; crochets in
a mesoseries
3. Body with 6-8 annulets per segment; A8 with a dorsal horn or button
3'. Body lacks annulets or has no more than six if present; dorsal horn on A8
absent4
4. Head sometimes retracted and hidden, body sluglike
 5. Head retracted and hidden; larva with spiny scoli, hairy tubercles or verrucae, rarely with gelatinous tubercles (some Asian species); crochets absentLIMACODIDAE The lack of crochets in limacodids distinguishes them from Lacturidae, Megalopygidae and Lycaenidae that share the same slug-like body form but have crochets on the prolegs on A3-6. Most identifications will be left at the family level. Distribution: worldwide. dorsal scoli white with red in middle of abdomen; Hawaii, various plants Darna pallivitta 5'. Head often retracted and hidden; crochets in mesoseries interrupted at center by spatulate lobe
Most lycaenids cannot be named past family unless they are from pineapple, pomegrantes, beans or New World cycads. Distribution: worldwide.

• honey gland present on A7; living larvae bright red; no modi	fied or star shaped
setae on body; from pineapple in Latin America	Strymon (see data sheet)
• pomegranates	possibly Virochola
New World cycads	
• beans, Mexicopossibly Every	es comyntas or Strymon melinus
• head not black, beans, Old World	•
• from <i>Pelargonium</i> , Europe or South Africa	1 1
6. Osmeterium present; the metathorax may be enlarged; transv	verse body stripes
and/or an eyespot may be present; some have fleshy filan	nentsPAPILIONIDAE
The osmeterium is not always everted in some preserved spe	
doubt, inject the larva with alcohol through the anus to try an	
or look for a slit on the prothoracic dorsum that is the exit ho	
osmeterium. Early instar papillionids often have short scoli a	
confused with Saturniidae. They are normally identified to fa	
are mature larva from Europe or Asia. Papilio demoleus is pr	
Caribbean Region and poses a threat to North America. The	<u>*</u>
Papilio demodocus, also from citrus, is almost morphologica	lly identical to
Papilio demoleus.	
• early instars with spiny scoli and a white mid-abdominal sade	•
larva smooth with a black or white intersegmental markings	
T3, another between A1 and A2, and a slanted lateral line on	
anterior margin of prothorax and A9 with paired small tubero	
abdominal segments never totally pure white; on citrus and r	
Caribbean	
6'. Osmeterium absent; metathorax and coloration not as above .	/
7. Abdominal segments two through six divided into five, or usua	ally six, annulets; crochets in
continuous, usually triordinal, mesoseries, sometimes with an ex	
parallel to the larger band	
There are two types of pierid larvae. One group has short secondary	
species have large chalazae mixed with smaller setae. Existing keys	
characters that rapidly lost in preserved larvae. Pest species feed on o	crucifers or legumes. Distribution:
worldwide.	
• on crucifersLeptophobia aripa, Pieris (bra	assicae, rapae) or Ascia monuste
• girdle present; transverse row of black dots on abdominal do	rsum; containers
from Europepo	ssibly pupae of Pieris brassicae
7'. Abdominal segments two through six not divided into annule	
lacking an extra short series of crochets parallel to the la	rger band8
8. Middorsal glands present A6 and A7	LYMANTRIINAE (EREBIDAE)
Although most lymantriines have glands on A6 and A7, one	
species apparent has lacks glands, a few species have only a	single gland
on A6, and in rare cases six gland slits may be present (Gard	ner 1941,
Kitching and Rawlins 1998). There are no morphological me	ethods to tell
gypsy moth strains apart but Wallner et al. (1994) suggested	discriminant

analyses of larval head colors. I do not know if this is practical. Pogue and Schaefer (2007) reviewed some selected Lymantria species and included a key to first instar larvae. Traxler (1977) studied the morphology of the European gypsy moth larva in detail with a scanning electron microscope. Pupae of Orgyia leucostigma plagiata can only be identified if the larval exuvium is present to determine head color. Distribution: worldwide. • head with two thick longitudinal stripes; dorsal verrucae spiny, without hairbrushes, tufts or flattened white setae; prothorax with a pair of lateral projecting verrucae; on trees or shrubs, usually not conifers; northeastern Pupa with dense dorsal abdominal setal tufts; the ground color sexually dimorphic (males light brown, females dark brown black); larval cast skin with the head brownish, not red or yellow, and with no trace of a setal tuft on A2 or red verrucae on the abdomen; origin has to be Nova Scotia and only from balsam fir around Christmas

8'. Middorsal glands absent A6 and A7.....9

9. Larva with barbed secondary setae and heteroideous crochets; if crochets are homoideous, then the mandible has a large molar lobeARCTIINAE (EREBIDAE)

Some Arctiinae have only a few scattered secondary setae on the L3 group of A1-8 that can be easily missed. Injecting the larvae, or boiling the specimen, helps to extend the prolegs making the crochets easier to see. The molar lobe of footman moths was illustrated by Stehr (1987). Species identification of *Utetheisa* depends on origin. Distribution: worldwide.

- head pattern with two black bands; setae yellow to brown to orange, on crops from Mexico and Central America often *Estigmene acrea*
- hairy pupae on bananas usually Ctenuchinae

10. Scoli absentVARIOUS FAMILIES

This is a collection of microlepidoptera and macrolepidoptera with secondary setae. Lecithocerid mandibular drawings show a large black tip; this may indicate they are sclerotized more heavily than normal, but this could not be evaluated without actual specimens. Pterophorid larvae on *Scabiosa* from Europe may be in the genus *Stenoptilia*, but no comparative material is available for comparison. *Sphenarches anisodactylus* is intercepted on pigeon pea from the Caribbean region. This couplet also includes noctuoids covered with secondary setae. As with the osmeterium, the prothoracic gland may not be visible in all preserved specimens; if it appears to be absent, double check by looking for a slit between the prothoracic legs. Any of these noctuids be could confused with Arctiinae. They are not frequently intercepted.

• mandible with a sclerotized tip on the first tooth; with verrucae and both sclerotized rings and a pore associated with SD1 on A8......Lecithoceridae

 anal point present; larva brown to gray to partially orange; with retractable tufts of steel blue to black hairs on T2 and T3; pine, Old World larvae covered with secondary setae that may be normal, spinelike, clubbed or curved; prespiracular pinaculum trisetose when secondary setae reduced; prolegs long and slender, crochets in uniordinal penellipse; cut flowers, 	
of steel blue to black hairs on T2 and T3; pine, Old World	
• larvae covered with secondary setae that may be normal, spinelike, clubbed or curved; prespiracular pinaculum trisetose when secondary setae reduced;	Dendrolimus
curved; prespiracular pinaculum trisetose when secondary setae reduced;	
Mexico, EuropeP	terophoridae
• prothoracic gland present; crochets uniordinal; prolegs well developed on A3-6	teroprioriane
	Pantheinae
• prolegs only on A4-6 N	•
• P2 high on head near vertex, distance between P2 less than distance separating	onaac (part)
P1; mandible of mature larva with no scissorial teeth; stipital lobe present; two	
MD setae present on A1	ontidae (part)
10'. Scoli present, either simple or branched	
10 . Scon present, either simple of branched	11
11. Stemmata not uniform in size or location; head angulate, sometimes with long	
scoli; middorsal scoli sometimes present on A7 but not A9; last abdominal	
segment may be forked; crochets uniordinal to triordinal, but usually	
triordinalN	VMDIIAI IDAE
Nymphalid larvae are highly diverse but APHIS rarely intercepts anything	IMPHALIDAE
• • • • • • • • • • • • • • • • • • • •	
except young larvae from herbaceous plants in Mexico. They are often	
confused with Saturniidae. It is likely they are all in the Nymphalinae.	
11'. Stemmata uniform in size and location; head smooth, without scoli;	
middorsal scoli sometimes present on A9 but not A7; last abdominal	
segment never forked; crochets biordinal	SATURNIIDAE
Saturniids are mostly intercepted as part of a Smuggling Interdiction and Trade	
Compliance (SITC) Program project. These usually involve eggs or pupae as	
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• prespiracular group fused with prothoracic shield; prothoracic spiracle, if oval,
• prespiracular group rused with promoracie sincid, promoracie spiracie, ir ovar,
is lying horizontal; SV group of T2 and T3 bisetose; case made mostly of plant
material such as leaves and twigsPsychidae
• larva often with six or less stemmata; three (rarely two) L setae on T1; L setae
on A1-8 widely separated; crochets may be surrounded by a band of cuticular
spines; case does not include much plant material if any at all
• aquatic larvae, gills present or absent, case of plant fragmentsAcentropinae
• from Chile on asparagus (not the true host), less commonly
intercepted in recent years; case smooth, not wrinkled, with a
triangular rear portion
• Hawaii possibly Hyposmocoma
 V1 setae on head macroscopic, half as long as P2; most often
cherry, apple or Euonymus, Canada and Europe
• large gregarious macrolepidoptera in a tent, pine or hardwoods;
• mandible with secondary setae, Old World
12'. Caterpillars not living in a case of plant or animal matter13
13. Caterpillars living in a leaf minevarious families
There are numerous characters on leaf mines that help identifications: the shape (blotch or
serpentine), the location (upper or underside) and the frass pattern if present. Thus, one should never
submit a leaf miner without the associated pressed mine. At least preserve the mine in the vial with the
larva. Good host data is also critical. APHIS rarely intercepts leaf miners, this seems strange, perhaps
inspectors don't recognize them. Or it is not properly noted on the APHIS form 309. Beside leaves,
mines also occur on bark and fruit. Pupal clusters of <i>Leucoptera malifoliella</i> are sporadically intercepted
from Europe or the Middle East, sometimes in large numbers. The tiny smooth white cocoons on apple
are often X-shaped. Gracillariids have hypermetamorphic development, there is a sap feeding early
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14'. Abdominal prolegs present on A615
15. Abdominal prolegs present only on A6 and A10 most GEOMETRIDAE
A few geometrids have vestigial abdominal prolegs on A3-5 or just A5. African
species can have well developed prolegs on A3-6. In spite of this being one of
the largest families of Lepidoptera, hardly any larvae go past family. A simple
rearing project would cheaply clarify the most common Mexican interceptions
from corn or raspberry. Specimens from Canadian blueberry sometimes can be
identified. Submit only mature larva with bright colors or atypical morphology.
There are no keys except for Canada and a picture book from Australia.
• Skin granulose; setae spatulate Sterrhinae (<i>Idaea</i> authority was cancelled)
15'. Prolegs present on other abdominal segments in addition to A6 and A1016
16. Four setae in the prespiracular group of the prothorax
APHIS has not yet intercepted <i>Langsdorfia</i> , but the larva will not key correctly
in any published work because of the extra seta on the L group of the
prothorax. It bores in stems through Latin America and is economically
important at times in Central and South America.
16'. Less than four setae in the prespiracular group of the prothorax17
10. Less than four setae in the prespiraction group of the promorax
17. Less than three setae in the prespiracular group of the prothorax18
17'. Three setae in the prespiracular group of the prothorax47
18. Apparently only one seta in the prespiracular group of the prothorax
Busseola does not occur in Europe. Setal angles are a key feature for African
stem borers so identification requires an eyepiece goniometer. This is a special
eyepiece for measuring angles like a protractor. APHIS has regularly
intercepted Sesamia from chestnuts even though it is not a recorded host of this
species.
• Europe, artichoke
• Africa, corn
• Europe, corn and chestnuts
18'. Two setae in the prespiracular group of the prothorax19
Most caterpillars intercepted by APHIS with a bisetose prespiracular group and
the L setae closely spaced belong to the Pyraloidea. More rarely they are
Carposinidae. However, when the world fauna is considered, a wide range of
other Lepidoptera have this chaetotaxy. Any of them could potentially be
confused with the Pyraloidea. Examples of a bisetose prespiracular group are
found in larvae of the Tineidae (Scardiinae), Gelechioidea (Momphidae)
Yponomeutoidea (almost all Glyphipterigidae, a few Yponomeutidae,
Ypsolophidae), Tortricidae, Copromorphoidea, Choreutidae (Millieriinae),
Epermeniidae, Alucitidae, Thyrididae, Hyblaeidae, Tineodidae, and
Brachodidae.
• Iris, Japan
• on Onagraceae, in stems or seedsprobably a true Momphidae

19. Setae L1 and L2 of abdominal segments 3 to 6 close together below the
spiracle, often on the same pinaculum
19'. Setae L1 and L2 of abdominal segments 3 to 6 widely separated, either both L
setae below the spiracle or one behind the spiracle (Noctuoidea, part)34
20. Four subventral setae present on abdominal segments 3 to 6CARPOSINIDAE
The submental flaps mentioned by Stehr (1987) are not obvious in the few
specimens of this family APHIS has intercepted.
• in fruit of <i>Malus</i> or <i>Crataegus</i> , Canada, Japan
20' Three subventral setae present on abdominal segments 3 to 6PYRALOIDEA
Compared to other large families of Lepidoptera (Noctuidae, Erebidae,
Tortricidae and Geometridae), the larger pyraloid subfamilies are relatively well defined and most authors use the same characters to define the
subfamilies. Because chaetotaxy of the Schoenobiinae is difficult to study, it is
easiest to look for their membranous sac, and if it is absent, eliminate this
subfamily as a choice before evaluating anything else.
With membranous sac anterior to prothoracic coxae; no sclerotized pinacula;
borers in rice or semi-aquatic plants; Latin America, AsiaSchoenobiinae
21 A
21. A sclerotized ring is present around at least SD1 on A8 or the L group of A9 is
trisetose (Pyralidae)
APHIS intercepts the Chrysauginae, Epipaschiinae, Galleriinae, Phycitinae and
Pyralinae. Sclerotized rings are sometimes hard to see. When in doubt, look for
a tiny sclerotized bar called a "neural connection" at the setal base or make a
slide of the cuticle and examine the pinacula with a compound microscope.
Either the sclerotized rings or L setae on A9 can be used to separate Pyralidae
from Crambidae. If the presence of rings is ambiguous, count the L setae. Distribution: worldwide.
• sclerotized rings on SD1 or D2 of the metathorax
• sclerotized rings on D2 of the mesothorax
21'. All segments lack a sclerotized ring and the L group on A9 is unisetose
(Crambidae)25
Some specimens of Pyraustinae and Spilomelinae have pigmentation on the
body pinacula that resembles a ring because the center is pale. It will be clear
that these are not the true sclerotized rings of the Pyralidae because of their
location and number. Those crambids with a bisetose L group on A9
(Acentropinae, Schoenobiinae) were discussed above. The organization of this
key by the reduced abdominal SD pinacula or the SV group on A1 is under
study. More obvious characters (extra pinacula present or prothoracic shield
pattern) are currently favored because identifiers will remember them easier.
22. Sclerotized ring present around SD1 of the mesothorax most Phycitinae
Phycitines have a sclerotized SD1 ring on the mesothorax, both the mesothorax
and A8, or on the mosothorax and at least partial rings on A1-8. More rarely
sclerotized rings are absent on all body segments (<i>Etiella</i> is a common
example). They can be either leaf rollers, stem borers, seed feeders or stored
product pests. Identification of other species of stored product pests are given
by Solis (2011) or Weisman (1987) and are not repeated here. Sibling species

- of *C. cactorum* occur in Peru and Argentina. *Hypsipyla robusta* occurs on Meliaceae in the Old World. Other species of *Etiella* are found in Asia. No morphological information useful for identification is present for any of these species. Distribution: worldwide.

- stored productspossibly another *Cadra* or *Ephestia* (includes *Anagasta*)

- prothoracic shield with characteristic pattern of two curved broken middorsal lines and a triangular or curved spot posterior to XD2; mandible with an inner tooth; SD1 seta of the mesothorax 1.5 times as long as the SD1 seta on the metathorax; D1 1/4 the length of D2 on A1; legumes, New WorldFundella pellucens
- hypopharyngeal complex with a series of blades; D2 posteroventrad of D1 on A1-7; body pinacula large and pigmented; SV group of A9 unisetose; Meliaceae, New World tropics
 Hypsipyla grandella
- early instar larvae are white; later instars with black head and prothoracic shield; orange to red larva transversely banded with black bands or spots; SV group bisetose on A7 and A8; *Opuntia*, New World, Australia, south Africa and Hawaiimature *Cactoblastis cactorum*

• mandible with an inner tooth on the first molar ridge and three scissorial teeth;
body dark; skin granulated; D setae of A1-7 in a horizontal line; L1 and L2 in a
vertical line on A3; corn and sorghum, Mexico, Central America
~
 prothoracic shield with a characteristic pattern of dark pigmentation, lightly
marked specimens have a single dark patch posterior to the SD1 seta, darkly
pigmented specimens have two such patches, one posterior to the SD1 seta, the
other ventrad of D2; no inner tooth on mandible; D2 posteroventrad of D1;
legumes, West Indies to Brazil
 cuticle dark and granulose with obvious tonofibrillary platelets; pineapple,
Cuba Ribua
22'. Sclerotized ring absent on SD1 of the mesothorax2
v > •••• v •=••• v •==• •• •=• •=• •=• •=•
22 Coloratized wing present around CD1 of A1 most Collowing
23. Sclerotized ring present around SD1 of A1 most Galleriina
Galleriinae divide up into several groups based on larval morphology.
Sometimes there are less than six stemmata and the SV group of T2 and T3 is
bisetose. Other genera have six stemmata and the SV group of T2 and T3 is
unisetose. The prespiracular group and prothoracic shield can be fused. A few
taxa have no sclerotized ring around SD1 of A1. Most interceptions are from
stored products or seed pods. However, the wax moth is sometimes imported as
pet food under permit requiring confirmation by port inspectors. Sclerotized
rings can be faint or obvious in Galleriinae.
• prespiracular and prothoracic shields fused; pineapple, Latin
America
 six stemmata present; peritreme of spiracles thickened on posterior margin;
sclerotized rings around seta SD1 on abdominal segments 1 and 8 not
complete; D1 and D2 pinacula on abdominal segments not pigmented; stored
products, cosmopolitan
<u>.</u>
• six stemmata present; peritreme of spiracles uniform in thickness;
sclerotized rings around seta SD1 on abdominal segments 1 and 8 complete;
D1 and D2 pinacula on abdominal segments pigmented; stored products,
cosmopolitanParalipsa gularis
• four stemmata present; SV group of T2 and T3 bisetose
 Prespiracular shield of prothorax extending below and behind the spiracle;
legume pods, tropical regions
23'. Sclerotized ring absent around SD1 of A12
24. Sclerotized rings absent on all body segmentsPhycitinae, Chrysaugina
• early instars have a solid black prothoracic shield; later instars have a
· ·
a characteristic pattern of four groups of dark spots on the prothoracic
shield, two lateral and two middorsal at the anterior and posterior
ends; legumes Etiella zinckenella (New World), Etiella sp. (Old World
• in Crescentia fruit, tropical America
24'. Sclerotized ring present on A8 Epipaschiinae, Pyralina
Most intercepted Pyralinae are stored products pests. There is no sclerotized
ring on A9 (misprint in some literature). Pyralis farinalis is the most common
species but scattered records from crops exist for P. manihotalis. There are no
morphological descriptions for <i>P. manihotalis</i> .

Epipaschiinae includes several pests of avocado but these are leaf feeders not likely to be seen because APHIS usually only imports only fruit. *Phidotricha* erigens is recorded from diverse plants but reared adults are most often from castor bean, corn and sorghum. *Pococera gelidalis* could be easily confused with *P. erigens* on *Mimosa pigra*, at least in Mexico and Honduras.

- First and sometimes the second molar ridge with an inner tooth; XD2 equidistant from SD1 and XD1; body with dark bands; tropical America castor
- Four stemmata; A9 with SV group uinsetose; abdomen light colored; stored
- Six stemmata: A9 with SV group bisetose; abdomen dark chocolate; stored

25. Mesothorax and metathorax with a single extra pinacula behind the dorsal setae or V1 of A9 twice as far apart as V1 on A10; most often stem borers in rice, reeds, corn, sorgum, sugarcane and grasses.......Crambinae

The pinaculum behind the dorsal setae on the mesothorax and metathorax is usually called an extra pinaculum. Other names are pinaculum without setae, non-setal bearing plate or secondary pinaculum. They may be faint or conspicuous. When these pinacula are absent or so faint that the larva must be cleared and slide mounted, it is better to stop at Crambinae unless the SV setal number or experience at the port suggests a better identification. Species identification of *Chilo* or *Diatraea* is very difficult and any pink or light colored stripes often fade rapidly in alcohol. Head and body color can be an important character, suggesting it is worth the effort to photograph larvae with a phone or cheap camera before preservation if a species name is critical. At the very least, when doing baggage interceptions from Mexico, be sure to get an exact locality for any corn or sugarcane *Diatraea*. The sclerotized patch of tonofibrillary platelets is anterior to the prothoracic coxae in many *Diatraea* was illustrated by Parada et al. (2007: fig. 7). Early instar Crambinae may be found on leaves or outside stems until they mature and become borers.

- prespiracular group not extending behind the prothoracic spiracle; SV group
- prespiracular group extending below and behind the prothoracic spiracle; SV group unisetose on mesothorax and metathorax; may have a tonofibrillary
- prothoracic shield has dark contrasting tonofibrillary platelets on anterior and posterior margin, those on the anal shield also dark and contrasting but with no obvious pattern; SV group bisetose on mesothorax and metathorax; no contrasting patch of tonofibrillary platelets anterior to the prothoracic coxae;

 • protnoracic shield tacks dark and contrasting tononormary platelets on both the anterior and posterior margin, anal shield with faint or no platelets, or if present, in a transverse line; SV group bisetose on mesothorax and metathorax; sometimes a sclerotized patch of tonofibrillary platelets is anterior to the prothoracic coxae; corn or sugarcane New World pest species of <i>Diatraea</i> (see data sheets and the Keys page on LepIn 25'. Mesothorax and metathorax with two extra pinacula behind the dorsal setae or extra pinacula absent; V1 on A10 twice as far apart as V1 on A9; larvae are borers or leaf rollers on a wide variety of hosts Extra pinacula are also found on the abdomen of some crambids (e.g. <i>Conogethes</i>). 	-
26. V1 pinaculum on A3-6 round; on Brassicaceae (Glapyriinae)	27
The latest concept of the Glapyriinae includes the Evergestiinae. Only a few of	
the potential pests APHIS could potentially intercept from this subfamily are	
included in this key. 26'. V1 pinaculum on A3-6 bandlike, rectangular or oval; on a wide range of hosts	
(Spilomelinae/Pyraustinae complex)	28
Not all members of the Spilomelinae/Pyraustinae complex have the bandlike or	
rectangular V pinacula on A3-6, but those that have these pinacula rounded do not feed on Brassicaceae.	
not reed on Brassicaceae.	
27. A tonofibrillary platelet in a sclerotized pit is present posterior to SD1 on A3-6; D1 longer than D2; D and SD pinacula not conical; prothoracic shield unmarked	Iellula
As with Crambinae, pink stripes can often fade rapidly in alcohol. It would be better to define <i>Hellula</i> with some morphological characters. The tonofibrillary platelet in a sclerotized pit posterior to SD1 on A3-6 is found in some <i>Crambus</i> , European Pyraustinae and <i>Hellula</i> . Its actual distribution or phylogenetic significance has not been studied. Usually D2 is longer than D1 in most Lepidoptera, <i>Hellula</i> is unusual in having the reverse. Orgin can help with species identification. <i>Hellula rogatalis</i> is North American; <i>H. undalis</i> is an Old World species and <i>H. phidilealis</i> occurs from the southern United States to parts of South America and Hawaii. <i>Crocidolomia pavonana</i> is just one of many pyraloids that may key out at this point so be careful of Asian interceptions on Brassicaceae.	· · · · · · · · · · · · · · · · · · ·
27'. No tonofibrillary platelet in a sclerotized pit present posterior to SD1 on A3- 6; D2 longer than D1; D and SD pinacula conical; prothoracic shield	
sometimes mottled	rgestis
There are several species of <i>Evergestis</i> that will key out here. <i>Evergestis rimosalis</i> from the New World has a mottled prothoracic shield, black conical D and SD pinacula and a characteristic pattern of cross stripes (hence the common name, cross striped cabbageworm). The D and SD pinacula of <i>E. forficalis</i> from Europe are conical but pale but only the SD pinacula are pigmented. There is no cross striped pattern and the SV group of T2 and T3 is bisetose.	J

28. Mesothorax and metathorax with a pair extra pinacula behind the dorsal
setaeSpilomelinae (part)
Several genera of Spilomelinae and have a pair of extra pinacula behind the
dorsal setae of the last two thoracic segments. Although literature sometimes
shows these pinacula on <i>Sceliodes</i> or <i>Pyrausta</i> , we could not confirm this in the
specimens we examined. Conogethes is a complex of species, our diagnosis is
based on specimens from Japan. Sometimes the two extra dorsal thoracic
pinacula of <i>Conogethes</i> almost fuse. The presence of a hairlike SD1 seta is
unusual.
 prespiracular group of the prothorax extends below and behind the
spiracle; A1-7 with one to three extra pinacula posterior to the spiracle;
sweet potato only
 prespiracular group of the prothorax does not extend behind spiracle;
mandible with two inner teeth and an outer "tooth" (lobe); front not
reaching the epicranial notch; SD1 pinacula of A2 and A7 not reduced;
no extra pinacula without setae on the abdomen; in pods or flowers of
legumes, tropical regions
 prespiracular group of the prothorax extends below and behind the spiracle;
extra pinacula on mesothorax and metathorax below L3; one extra pinacula
behind the L1 and L2 setae on A1-7; SD1 on A9 hairlike; young larvae with no
pinacula; traditionally from peach, pine, castor bean, Asia
28'. Mesothorax and metathorax lack an pair extra pinacula behind the dorsal
setae29
29. SV group of A1 unisetose on at least one side; D1 pinacula of A2-8
often contains a pale or dark spot; only on Solanaceae
(Spilomelinae, part)
Neoleucinodes and Leucinodes. The SV group of A1 is unstable, but usually
unisetose. It is better to trust the origins over morphology if the identification is
in doubt
• body pinacula pigmented, S1 posterior of a line connecting stemmata 2 and 3;
Old World
 stemma 1 and stemma 2 closely spaced; S1 is touching or anterior to a
 stemma 1 and stemma 2 closely spaced; S1 is touching or anterior to a vertical line connecting stemmata 2 and 3; prothoracic shield without
 stemma 1 and stemma 2 closely spaced; S1 is touching or anterior to a vertical line connecting stemmata 2 and 3; prothoracic shield without spots or faintly mottled at most; body pinacula pale; SD1 pinacula on A2
 stemma 1 and stemma 2 closely spaced; S1 is touching or anterior to a vertical line connecting stemmata 2 and 3; prothoracic shield without spots or faintly mottled at most; body pinacula pale; SD1 pinacula on A2 and A7 not reduced; SD2 seta and pinaculum on A3-6 inconspicuous;
 stemma 1 and stemma 2 closely spaced; S1 is touching or anterior to a vertical line connecting stemmata 2 and 3; prothoracic shield without spots or faintly mottled at most; body pinacula pale; SD1 pinacula on A2 and A7 not reduced; SD2 seta and pinaculum on A3-6 inconspicuous; SD1 anterior or anterodorsad of the spiracle on A8 and the host is a
• stemma 1 and stemma 2 closely spaced; S1 is touching or anterior to a vertical line connecting stemmata 2 and 3; prothoracic shield without spots or faintly mottled at most; body pinacula pale; SD1 pinacula on A2 and A7 not reduced; SD2 seta and pinaculum on A3-6 inconspicuous; SD1 anterior or anterodorsad of the spiracle on A8 and the host is a cultivated species of <i>Solanum</i>
 stemma 1 and stemma 2 closely spaced; S1 is touching or anterior to a vertical line connecting stemmata 2 and 3; prothoracic shield without spots or faintly mottled at most; body pinacula pale; SD1 pinacula on A2 and A7 not reduced; SD2 seta and pinaculum on A3-6 inconspicuous; SD1 anterior or anterodorsad of the spiracle on A8 and the host is a
 stemma 1 and stemma 2 closely spaced; S1 is touching or anterior to a vertical line connecting stemmata 2 and 3; prothoracic shield without spots or faintly mottled at most; body pinacula pale; SD1 pinacula on A2 and A7 not reduced; SD2 seta and pinaculum on A3-6 inconspicuous; SD1 anterior or anterodorsad of the spiracle on A8 and the host is a cultivated species of Solanum
• stemma 1 and stemma 2 closely spaced; S1 is touching or anterior to a vertical line connecting stemmata 2 and 3; prothoracic shield without spots or faintly mottled at most; body pinacula pale; SD1 pinacula on A2 and A7 not reduced; SD2 seta and pinaculum on A3-6 inconspicuous; SD1 anterior or anterodorsad of the spiracle on A8 and the host is a cultivated species of <i>Solanum</i>
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 stemma 1 and stemma 2 closely spaced; S1 is touching or anterior to a vertical line connecting stemmata 2 and 3; prothoracic shield without spots or faintly mottled at most; body pinacula pale; SD1 pinacula on A2 and A7 not reduced; SD2 seta and pinaculum on A3-6 inconspicuous; SD1 anterior or anterodorsad of the spiracle on A8 and the host is a cultivated species of Solanum

them) is not clear. None of the three species below have SD1 reduced on A2 or A7 except <i>O. indicata</i>	
• prespiracular group of the prothorax on a nearly crescent-shaped pinaculum;	
prothoracic shield with a dark and sharply defined spot; D setae of mesothorax	
on the same pinaculum; the top mesothoracic SD seta is anterior of the bottom	
one; body pinacula unpigmented; SV group is bisetose on A1; crochets in a	
complete or nearly complete biordinal circle; Solanaceae, New WorldLineodes integra	
• prothoracic shield mottled with a pair of black dots; D setae of mesothorax on	
the same pinaculum; the top mesothoracic SD seta is posterior of the bottom	
one; SD pinaculum of the mesothorax, and sometimes the metathorax,	
pigmented; SV group of A1 bisetose; herbaceous crops and weeds,	
cosmopolitan	
• front extends less than 1/2 the distance to the epicranial notch; mandible	
triangular in shape and usually without teeth (one to three small teeth may be	
present); SV group bisetose on A1; legumes, subtropical regions	
• prespiracular pinaculum of the prothorax oval; D setae of mesothorax on	
separate pinaculum; the top mesothoracic SD seta is posterior of the bottom	
one; crochets in a complete circle, triordinal mesally and uniordinal laterally;	
polyphagous, cosmopolitan	21
This group includes two species with a partially dark prothoracic shield. As pointed out by Hayden et al. (2013), <i>R. periusalis</i> is another example using red/pink coloration for identification is limited because this pigment fades in preserved pyraloid larvae. • prothoracic shield shaded with black almost to the D1 seta; SV group of A1 bisetose; D1 pinaculum on the abdominal segments with a black spot anterodorsad to the D1 seta; D2 pinacula on A8 not fused; southern U.S. to Brazil, Solanaceae	
prothoracic shield is patterned differently or is completely dark	32
32. Mesothoracic D and SD pinacula fused	art)
This group includes species of <i>Herpetogramma</i> with a pale prothoracic shield.	
Two of them, Herpetogramma licarsisalis and Herpetogramma phaeopteralis,	
are pests of grasses. Not all early instars of at least some species have the	
mesothoracic D and SD pinacula fused (Passoa 1985: 112).	22
32'. Mesothoracic D and SD pinacula separate	33

33. Head capsule with a shield like extension over base of antenna; or prespiracular
group of the prothorax extends below and behind spiracle to fuse with the
posteroventral corner of the prothoracic shield; or a tonofibrillary platelet in
a sclerotized pit is present posterior to SD1 on A3-6 misc. Pyraustinae
This couplet separates a few miscellaneous Pyraustinae from the remaining
Spilomelinae. <i>Ostrinia</i> is a complex of species that is especially hard to identify
to species outside North America. The extension of the prothoracic shield in
Achyra is sometimes lightly sclerotized and hard to see. Identification of
Pyrausta is restricted to plants in the Lamiaceae pending a better larval
characterization of the genus.
• head capsule with a shield like extension over base of antenna, wide
range of herbaceous hosts, especially corn and
pepper
• prespiracular group of the prothorax extends below and behind
spiracte to fuse with the posteroventral corner of the
prothoracic shield
• mesothorax and metathorax with the MD and MSD setae on pigmented
1 6
pinacula; SD1 pinacula of A2 and A7 not reduced; a tonofibrillary platelet in a
sclerotized pit is present posterior to SD1 on A3-6; Canada, Europe, on
Lamiaceae — Pyrausta
33'. Head capsule lacks a shield like extension over base of antenna and the
prespiracular group of the prothorax does not fuse with the prothoracic
shield and a tonofibrillary platelet in a sclerotized pit is absent posterior to
SD1 on A3-6(Spilomelinae, Cybalomiinae, part
This group includes miscellaneous species of Spilomelinae, some of which are
frequently intercepted. Hendecasis duplifascialis, a member of the
Cybalomiinae, also keys out here. <i>Nomophila</i> is a complex of species that,
except for Europe and North America, have poorly known immatures. The
same is true for <i>Diaphania</i> in Latin America.
 XD2 equidistant from SD1 and XD1; thoracic pinacula not darkly pigmented;
SV group on A1 trisetose; crochets of A3-6 biordinal; bores in buds of jasmine
from Asia
 XD2 closer to SD1 than XD1; one SD seta on T2 and T3 hairlike; SV pinacula
of T2 and T3 notched anteriorly; SV group trisetose on A1; SD pinacula
reduced on A2 and A7; anal shield lacks pigmented patches; polyphagous on
herbaceous plants but especially on peppers, cut flowers, sometimes
saprophagous, southern Europe, Middle East and AfricaDuponchelia fovealis
VDQ 1 (CD14 VD1 CV) A1 CD 1 1
• XD2 closer to SD1 than XD1; SV group trisetose on A1; SD pinacula
• XD2 closer to SD1 than XD1; SV group trisetose on A1; SD pinacula reduced on A2 and A7; anal shield with pigmented patches; polyphagous
reduced on A2 and A7; anal shield with pigmented patches; polyphagous
reduced on A2 and A7; anal shield with pigmented patches; polyphagous on herbaceous plants
reduced on A2 and A7; anal shield with pigmented patches; polyphagous on herbaceous plants
reduced on A2 and A7; anal shield with pigmented patches; polyphagous on herbaceous plants
reduced on A2 and A7; anal shield with pigmented patches; polyphagous on herbaceous plants
reduced on A2 and A7; anal shield with pigmented patches; polyphagous on herbaceous plants
reduced on A2 and A7; anal shield with pigmented patches; polyphagous on herbaceous plants

Species identification should not be attempted unless the specimen is from India or Central Africa on cotton. *Earias* is one of the most frequently misidentified caterpillars until the new identifier learns the characters and host/orgin.

34'. Dorsum of larvae without spiny fleshy filaments; SV group of thorax variable......35

Noctuoid larval family identification is at best challenging. For the moment, an attempt is made to at least mention all the taxa as painfully unsatisfactory it may be to characterize them. Without an ability to separate these families, the most accurate approach is to drop back to Noctuiodea instead of guessing at a probable answer. Unfortunately, this would hurt the resolution of the Pest ID database and make it pointless to consult. Practically speaking, APHIS has to make an educated guess and live with wrong answers at times. When the larva is from Anacardiaceae, and the crochets are heteroideous, the larva is most likely to be in the Eutellidae. Oenosandridae are rare and distinctive. All Notodontidae are difficult to separate from all Noctuidae. No single character defines all larval Nolidae. There is no set of characters to separate all Noctuidae from all Erebidae. Separation of these two families is discussed in the "escape couplets". Identifiers should expect numerous exceptions although many common species will key out properly.

- P2 high on head near vertex, distance between P2 less than distance separating P1; mandible of mature larva with no scissorial teeth; stipital lobe present; two MD setae present on A1; seta X (or verruca) present near anterolateral corner of anal shield; maxillae with stipital lobes; head is often striped with two closely spaced parallel bands of different colors and the vertex may be pointed; fleshy humps on the abdominal dorsum or a small knob on A8 may be present;

- CC 411144	rect
off the substrateN	`L '
• dorsum with very long setae; F setae well above frontal pores	possibly Nolidae
35. Prolegs reduced or absent on A3 and/or A4 compared to A5 and A6	
(Noctuidae/Erebidae complex, part)	
For nearly 100 years the Noctuidae adult classification by Hampson was ri	gidly
followed without regard to the input of larval characters. Now molecular	
studies change the classification immediately. As a result, there is no easy	to
use larval subfamily key for Noctuidae or Erebidae, and for most tropical	
regions, there is nothing comprehensive for species identification. Dissecti	
the mouthparts is needed for many groups. Realistically, it is impossible to	
identify early or middle instars, which unfortunately is the bulk of what Al	
intercepts in Noctuoidea. Given these constraints, the goal of APHIS shoul	
to name the pests that represent the most serious risk and separate them from	
species we have in the US, while identifying the rest of the unknowns to the	e
family level unless they are unusual. That is the goal for this section.	
35'. Prolegs of A3-A6 all equal in size	
Be sure to check the anterior prolegs carefully because in some Erebinae the	
reduction is slight. Early instars of some subfamilies (e.g. Noctuinae) have	
reduced prolegs but in later instars the proleg size becomes more equal.	
Therefore, this key will not work with early instars if the proleg size is red	
These are best identified only to family or Erebidae/Noctuidae unless they	are
Plusiinae which can be recognized by their biordinal crochets.	
36. Seta V1 on A1 and A2 modified into a ringlike structure	Erebinae (part)
	_
This taxa was named Group I of the Catocalinae by Crumb (1956). Melipo	tis is
the most commonly intercepted genus. Be sure to check for the ringlike ser	ta on
the most commonly intercepted genus. Be sure to check for the ringlike set both A1 and A2. If four rings are seen, you can be sure this modification is	ta on
the most commonly intercepted genus. Be sure to check for the ringlike ser	ta on
the most commonly intercepted genus. Be sure to check for the ringlike set both A1 and A2. If four rings are seen, you can be sure this modification is present. Setae that fall out of their socket are easily confused with the ringli setae.	ta on s ike
the most commonly intercepted genus. Be sure to check for the ringlike set both A1 and A2. If four rings are seen, you can be sure this modification is present. Setae that fall out of their socket are easily confused with the ringle	ta on s ike
the most commonly intercepted genus. Be sure to check for the ringlike set both A1 and A2. If four rings are seen, you can be sure this modification is present. Setae that fall out of their socket are easily confused with the ringli setae.	ta on sike37
the most commonly intercepted genus. Be sure to check for the ringlike set both A1 and A2. If four rings are seen, you can be sure this modification is present. Setae that fall out of their socket are easily confused with the ringle setae. 36'. Seta V1 on A1 and A2 minute but normal in form, not ringlike	ta on sike37Plusiinae
the most commonly intercepted genus. Be sure to check for the ringlike set both A1 and A2. If four rings are seen, you can be sure this modification is present. Setae that fall out of their socket are easily confused with the ringle setae. 36'. Seta V1 on A1 and A2 minute but normal in form, not ringlike	ta on sike37Plusiinae si but
the most commonly intercepted genus. Be sure to check for the ringlike set both A1 and A2. If four rings are seen, you can be sure this modification is present. Setae that fall out of their socket are easily confused with the ringle setae. 36'. Seta V1 on A1 and A2 minute but normal in form, not ringlike	ta on sike37Plusiinae si but
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the most commonly intercepted genus. Be sure to check for the ringlike set both A1 and A2. If four rings are seen, you can be sure this modification is present. Setae that fall out of their socket are easily confused with the ringle setae. 36'. Seta V1 on A1 and A2 minute but normal in form, not ringlike	and to
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the most commonly intercepted genus. Be sure to check for the ringlike see both A1 and A2. If four rings are seen, you can be sure this modification is present. Setae that fall out of their socket are easily confused with the ringle setae. 36'. Seta V1 on A1 and A2 minute but normal in form, not ringlike	Ta on sike
the most commonly intercepted genus. Be sure to check for the ringlike set both A1 and A2. If four rings are seen, you can be sure this modification is present. Setae that fall out of their socket are easily confused with the ringl setae. 36'. Seta V1 on A1 and A2 minute but normal in form, not ringlike	Ta on sike

• vestigial prolegs present on A3 and A4; SD1 hairlike on A9; molar ridges continue to the scissorial teeth; D and SD setae equally spaced on T2 and T3: texture of cuticle smooth to slightly granular depending on the magnification; ten ridges present on the raduloid; highly polyphagous, New World, Hawaii, North Africa, Middle • head with setal bases dark, genal spot usually present; subdorsal area with pigmented SD1 pinacula and a white stripe in living larvae; thoracic legs black or green; vestigial prolegs present on abdominal segments 3 and 4; SD1 hairlike on A9; mesothoracic D setae almost touch; two molar ridges do not reach the scissorial teeth; pinacula not surrounded by ring of minute black spicules; polyphagous • Hawaii Chrysodeixis eriosoma • head with setal bases dark, genal spot usually present; subdorsal area with pigmented SD1 pinacula and a white stripe in living larvae; thoracic legs black or green; vestigial prolegs present on abdominal segments 3 and 4; SD1 hairlike on A9; diameter of pigmented area on SD1 pinacula of A1-7 equal to or larger than the diameter of the spiracle; sometimes with thoracic legs and all body pinacula dark; raduloid with 10 ridges; polyphagous, North America • Vestigial prolegs present on abdominal segments 3 and 4; SD1 hairlike on A9; setal bases of head and epicranium similar in color; with 23 ridges on raduloid; polyphagous, subtropical and tropical regions of New • prolegs completely absent on abdominal segments 3 and 4; SD1 on A9 thin but setaform; A2 with the SV1 and SV2 pinacula separated; ventral setae surrounded by a ring of black spicules; mandible ribs continue to cutting • prolegs completely absent on abdominal segments 3 and 4; SD1 on A9 thin but setaform; pinacula of setae SV1 and SV2 fused on abdominal segments 2 to 4; adfrontal area with thin oblique vertical dash; genal dash present or absent; inner face of mandible black in late instars, most abdominal setae pale in late instars; minute ring of microspines surround lower body pinacula, most pronounced in early instars.... • prolegs completely absent on abdominal segments 3 and 4; SD1 on A9 thin but setaform; A1 with bisetose SV group; A2 with the SV1 and SV2 pinacula at least partially fused; cuticle covered with long spinules, the dorsal ones white and the lateral ones black; ventral abdominal pinacula not ringed with black spicules; polyphagous, North and Central America to Costa Rica Megalographa biloba

• prolegs completely absent on abdominal segments 3 and 4; SD1 on A9 thin but setaform; A1 with bisetose SV group; A2 with the SV1 and SV2 pinacula at least partially fused; cuticle covered minute granules; ventral pinacula ringed with small black spinules; polyphagous, western United States to Baja	
California and maybe northern Mexico	
37'. Crochets clearly uniordinal and uniserial miscellaneous Erebidae	e/Noctuidae
This couplet includes the loopers and semiloopers that are not Plusiinae. The	
most likely subfamilies are Acontiinae (Noctuidae) and Erebinae (Erebidae).	
Achaea janata is common in tropical regions and was intercepted on Euphorbia	
from Hawaii. <i>Plusia</i> also keys out here because it has uniordinal crochets, an	
exception for the subfamily. <i>Anticarsia gemmatalis</i> , which feeds on legumes	
and especially soybean, would key out here.	
38. Cuticle rough and spiny	39
Use about 100 to 200 power on a stereoscope to evaluate the cuticle texture. A	
very common mistake is to assume all noctuids with a spiny cuticle must be in	
the Heliothinae. In fact, both the Erebidae and Noctuidae contain species with a	
spiny cuticle.	
38'. Cuticle smooth or granular, not spiny	40
39. Prothoracic L setae arranged horizontally or in a slanted line; more often feeding on fruits or flowers than leaveslate instar The horizontal arrangement of the prothoracic setae is a distinctive character of	Heliothinae
the Heliothinae. Unfortunately, it is not present in the early instars. Early instar	
Heliothinae often look different from the mature larva as well.	
There are no known characters to separate Helicoverpa armigera from	
Helicoverpa zea in Brazil. Identification of H. armigera in the Old World is	
equally difficult. The guide below for <i>H. armigera</i> , assulta and puntigera is to	
help select samples for barcoding more than for making quarantine decisions.	
A more detailed key for <i>armigera</i> suspects is available on the Keys page of	
LepIntercept, but in spite of being more detailed, it is still provisional. Like the	
Plusiinae, APHIS wastes much time and effort cataloging pointless repeated	
interceptions of the Central American species on the same crops year after	
year. Each one requires mandible dissection or looking at the pinacula under	
high power. Worse yet, unknown <i>H. armigera</i> infestations in the New World	
would be identified as <i>H. zea</i> resulting in specialists screening for a target that	
will never be recognized with morphology. A general Heliothinae key is available on the Keys page on LepIntercept, but the document badly needs to	
be updated and hopefully will be the subject of its own identification tool in the	
future. For now, a better policy is to identify all New World Heliothinae to	
subfamily except for <i>H. zea</i> that could be randomly barcoded. All Brazilian <i>H</i> .	
zea interceptions should be considered H. armigera suspects.	
• D setae of A1-8 inserted on large conical chalazae, those of A1, A2 or A8 often	
larger than the rest; body color highly variable, but usually with lines and	
stripes and sometimes a black bar joining the D setae of A1 or A2 (saddle	
marking); if the setal bases are small, then the mandible has a minute tooth on	
the inner rib and no large retinaculum (<i>H. armigera</i> suspect, see data sheet)	

 microspines uniformly covering body; Old World, on
Solanaceaeusually <i>H. assulta</i>
• three rows of body microspines; spiracles brown; prothoracic
setae dark; no saddle marking on A1 or A2; polyphagous,
Australia probably H. puntigera
 three rows of body microspines; spiracles black or brown;
prothoracic setae pale; with black bar (saddle marking)
connecting the D setae on A1 or A2; polyphagous, Old World
(sibling species in Africa), Brazil probably H. armigera
 mandible with a large retinaculum, sometimes reduced to a thin ridge or
groove; at least half of the dorsal pinacula of A1, A2 and A8 covered with
microspines; conical pinacula, if present, are only on A1, A2 and A8; color and
pattern highly variable; host range wide,
Central America, Caribbean, Guyana, Suriname, French Guiana
and Hawaii
• rest of South America
• Chile Heliothinae
• at least half of the dorsal pinacula of A1, A2 and A8 covered with microspines;
SD1 and L1 setal bases of A4 connected to each other by a band of microspines (sometimes barely so) and both are much larger than the diameter of the
spiracle; feeds primarily on <i>Physalis</i> , only rarely on <i>Solanum</i>
• mandible lacks a retinaculum or at most has a trace of a small tooth; dorsal
pinacula of A1, A2 and A8 lacks microspines or have only a few around the
edges; conical pinacula, if present, are only on A1, A2 and A8; color and
pattern highly variable; host range wide but generally not associated with
Physalis,
 Central America, Caribbean, Ecuador, Colombia, Venezuela and
Hawaii
• rest of South America(Helicoverpa sp.)
39'. Prothoracic L setae arranged verticallyvarious Noctuidae/Erebidae
This is the common condition for most noctuids and seems not to vary by
instar. Below are three examples of non-Heliothinae taxa with a spiny cuticle
that are occasionally intercepted. Plusiinae also have a spiny cuticle, but they
are easily recognized to subfamily using the characters noted above. The spiny cutworm from Chile has been called " <i>Agrotis lutescens</i> " or <i>Feltia malfida</i> but it
needs to reared and carefully identified.
• SD1 hairlike on A9; Chile only
• at least dorsal setae stout and faintly clubbed; cosmopolitan
 head often large for size of body, may be bicolored; prothoracic SV group
trisetose; body pinacula conical, often with a pale center giving the impression
of "rings"; some dorsal or subdorsal setae very long; prepupa pale; various
plants, MexicoLitoprosopus
40. Head with a characteristic thick longitudinal band; dorsal spots on A2
usually present; pineapple, Central America and northern South
AmericaElaphria ''nucicolora''

The pupa and larva of <i>Elaphria nucicolora</i> are frequently intercepted in
association with pineapple. Although there is some larval color variation, a few
rearings and dissected genitalia from pupae have always been this species.
Literature and web photos show a thin light middorsal stripe. The intercepted
Elaphria fauna needs study; quotes are used around the species name to
suggest further confirmation.
arva lacks both a thick longitudinal band and dorsal spots on A2; host and
origins vary
Many nostyid/arabid larges have a thick head hand but these look spots on A?

40'. L	Larva lacks both a thick longitudinal band and dorsal spots on A2; host and origins vary	41
	Many noctuid/erebid larvae have a thick head band but these lack spots on A2 and/or an association with pineapple.	
41. M	Iesothorax and metathorax lack a minute tonofibrillary platelet connected to the SD setae by a sclerotized bar	42
41'. N	Mesothorax and metathorax with at least SD1 (and sometimes SD2 as well)	40
	connected to a minute tonofibrillary platelet by a sclerotized bar	43

Feltia subterranea is the most easily recognized member of this group. Some Agrotis ipsilon can be identified from Europe or Canada. Other unknown species are best left at genus or subfamily. Most of these larvae tend to avoid woody plants. The identification is more accurate if the host is an economically important crop.

the below may help in special cases.

These larvae may possess one or more, but not all, of the characters defining the *Agrotis/Feltia* complex. After the Plusiinae are eliminated as a possibility, if desired, a port identifier can make an educated guess and put the results in the comment section of PPQ form 309. Do not forward difficult examples (SD1 of intermediate thickness) or poorly preserved specimens to specialists. APHIS gets too many noctuid/erebid interceptions to spend hours with each find, but

- spinneret with silk pore concealed by apical flaps (illustrated by Crumb 1956: plate 3 I-R); SV group trisetose on A1 and SD1 setaform on A9possibly Erebidae
- spinneret with the silk pore open apically (illustrated by Crumb 1956: plate 2); SV group bisetose on A1 and SD1 setaform on A9possibly Noctuidae

43. H	ead with adfrontal area outlined in white forming an inverted "Y"; mandible with four scissorial teeth and no retinaculum; SD1 on T2 and T3
	connected to the associated tonofibrillary platelet by a minute sclerotized bar; SV group bisetose on A1; lateral spot often present on the mesothorax
	or A1 and body setae short, most not much longer than the vertical height
	of the spiracle on A8Spodoptera spp.
	Larval <i>Spodoptera</i> are hard to define morphologically but they do present a
	characteristic appearance. Many Latin American pests also occur in the United
	States. Consult Passoa (1991, 2011) for New World species not mentioned
	below. The key by Pogue (2002) has the advantage of using preserved larvae
	without a need for origin. Two Old World species (S. litura and S. littoralis)
	are quarantine significant and very variable in color. The <i>Spodoptera</i> key on
	the Keys page on LepIntercept gives guidelines for their separation, but it is
	not possible to correctly identify all the atypical color forms. Origin helps as
	the two are mostly allopatric. Young larvae cannot go past genus except for S.
	litura on orchids from Thailand.
	• small larvae defined by the swollen thorax
	• orchids, Thailand
	• ground color green to yellow brown to dark blue gray; subdorsal area often not
	contrasting with paler dorsum; middorsal line often obvious; spiracular stripe
	often interrupted on A1 by a black band or spot; dorsal triangles, if present, are
	on all abdominal segments, A1 and A8, A7 and A8 or just A8 and most of
	them usually have an apical white dot; abdominal spiracles usually with a large
	black dot dorsally and a white spot posteriorly; from Middle East to Asia on a
	wide range of hosts
	 ground color a shade of chocolate brown to steel gray to dark olive green;
	subdorsal area usually strongly contrasting with paler dorsum; middorsal line
	usually faint or absent; spiracular stripe not interrupted on A1 by a black band
	or spot; dorsal triangles, if present, are on all abdominal segments, A1 and A8,
	A7 and A8 or just A8 and in most cases lack an apical white dot; sometimes a
	white spot is present posterior to the abdominal spiracles, more rarely with a
	dorsal black dot; from Europe to Africa to the Middle East on a wide range of
	hosts
	 mesothoracic lateral dark spot normally present near SD1; large lateral spot on
	A1 absent; dorsum often contrasting with the darker subdorsal area and colored
	with an irregular series of white dots and broken lines, more rarely a series of
	paired thin black dashes are present; cosmopolitan, polyphagousS. exigua (see data sheet)
	 Dorsal abdominal pinacula larger than diameter of spiracles on A1-7, these
	pinacula either conspicuous, especially on A8 and A9 (brown or black color
	forms) or pale (green form); abdominal segments with a granulated texture
	under a magnification of 25x or greater
43'. I	Mesothorax and metathorax with both SD1 and SD2 connected to a minute
	tonofibrillary platelet by a sclerotized bar; adfrontal area of head often not
	white; larva lacks a swollen thorax or lateral dark spot on the mesothorax
	or A144

44. Spinneret with apical spinules; spiracles black; four to eight yellow middorsal
spots usually present, sometimes also with a W-shaped mark on A8;
polyphagous, cosmopolitanPeridroma saucia
The presence of yellow middorsal spots is diagnostic for <i>P. saucia</i> but they can
be faint or absent. This species often occurs with <i>Copitarsia</i> and would be a
<u>*</u>
common non-target in <i>Copitarsia</i> larval surveys. Color forms of <i>P. saucia</i>
without any dorsal spots will be hard to identify without dissecting the
mouthparts. There are sibling species in Chile, Argentina and Hawaii.
44'. Spinneret lacks apical spinules; spiracle color variable; yellow middorsal
spots absent; W-shaped mark sometime present on A845
45. Spinneret rounded or with a medial depression; labial palpi with last segment much shorter than basal segment; mature larvae with white body setae, paired dorsal dashes or triangles, lateral red spots, or no markings; young larvae with a mottled head, dark setae, reduced prolegs on A3 and A4 and a green dorsum faintly striped with white; cut flowers and vegetables most often from Mexico, northern South America and Chile, rarer in Central America except for Guatemala
Mexico and Central America
45'. Mandible and spinneret variable; labial palpi with last two segments longer
than one third the length of the basal segment; coloration, hosts and
origins variable46
01151115 74114010
46. Mandible with a large retinaculum; spinneret about four times longer than wide; last segment of labial palpi about as long as the basal segment; sixteen stout spinelike blades present on the hypopharyngeal complex; abdominal prolegs with a lateral sclerotized plate; normal (dark) form with a black patch surrounding spiracles of A1-8; green form with thick white spiracular stripe extending from the head to the anal prolegs, passing through the spiracles of A1-7 but below the larger spiracle of A8; polyphagous, Old World origins only

46'. No black patch surrounds the spiracles of A1-8 or no thick white spiracular
stripe from the head to the anal prolegs is present; if this coloration is seen,
then mouthparts not as above; worldwide on many hosts Noctuida
This couplet was added so that unknown larvae will not be forced into
Copitarsia or M. brassicae because the key "dead-ends" into these two choices.
As in couplet 42, the same choices and guidelines apply.
• spinneret with silk pore concealed by apical flaps (illustrated by Crumb 1956:
plate 3 I-R); SV group trisetose on A1 and SD1 setaform on A9possibly Erebidae
• spinneret with the silk pore open apically (illustrated by Crumb 1956: plate 2);
SV group bisetose on A1 and SD1 setaform on A9possibly Noctuidae
• spinneret with the silk pore dorsally grooved, wider than long or fringed
(illustrated by Crumb 1956: plate 2); SV group bisetose on A1 and SD1
hairlike on A9 possibly Noctuinae (including Hadeninae)
namike on A3possiony Noctumae (including fladenmae)
47. D2 of A9 on a single pinaculum, or if on separate pinacula, then the D2 setae
are closer to each other than each D2 seta is to D1 below it; D1 and SD1 of
A9 sometimes on the same pinaculum; SD1 on A9 never hairlike; anal
comb, if present, with straight teethTortricida
Currently, the family Tortricidae includes three subfamilies: Tortricinae,
Olethreutinae, and Chlidanotinae. Among the Tortricinae, most pest species are
in the Archipini, usually as external feeders (leaf rollers). The Olethreutinae
contain many pest species in the Grapholitini that feed internally in fruits or
stems. Chlidanotinae larvae are poorly known; most bore in twigs, fruits or
seeds. The former family "Cochylidae" is a tribe of the Tortricinae (Cochylini)
and is no longer recognized as a separate family.
Larval tortricids are normally recognized by having the prespiracular group
trisetose (but see couplet 19), L1 and L2 on A1-8 closely spaced, the abdominal
crochets in a circle or ellipse, and the D2 setae on A9 joined on the same
pinaculum (called a saddle pinaculum by Gilligan and Epstein 2012). When D2
are on separate pinacula, or the pinacula are faint, tortricids are recognized by
the D2 setae being more closely spaced to each other than to their
corresponding D1 setae. Only a few tortricids have the D2 setae widely spaced.
SD1 on A9 is never hairlike in Tortricidae, although this character state is
commonly seen in Gelechioidea. Most Olethreutinae, and some Tortricinae,
have D1 and SD1 of A9 joined on the same pinaculum. This is an unusual
arrangement as well. Unfortunately, numerous exceptions prevent reliable
separation of tortricid subfamilies in the larval stage. A better approach is to
classify them into phenetic groups or "types" as proposed by Brown (2011). A
modified version of his key by Todd Gilligan is available on the Keys page on
LepIntercept.
47'. D2 of A9 on separate pinacula or each D2 seta closer to their corresponding
D1 seta than to the other D2; D1 and SD1 of A9 not joined on the same
pinaculum; SD1 on A9 sometimes hairlike; anal comb often absent, if
present, teeth are straight or curved4
This couplet leads to those frequently intercepted microlepidoptera with a
trisetose prespiracular group (Gelechioidea, Yponomeutoidea and Tineidae).
The anal comb is generally absent except for Gelechiidae and some other rarely
seen Gelechioidea. Gelechioids have straight or curved teeth whereas the teeth

are always straight in Tortricidae. A few microlepidoptera (Choreutidae and Gelechioidea) have the D2 setae joined on A9 but they differ from Tortricidae in other ways.

48. Sul	bmentum with a large oval pit; A1-8 often with a sclerotized ring around
	SD1; live or decayed plant material, cosmopolitan
	BLASTOBASIDAE, OECOPHORINAE (Endrosis)
	Two generalizations on blastobasids require modification. First, there are other
	Gelechioidea with submental pits or similar structures. One source of confusion
	is with Oecophorinae, particularly those found under bark. Another is with
	Endrosis from stored products. Xyloryctidae from Australia have a pit although
	it is not clear if this structure is homologous with the Blastobasidae.
	The second generalization is that blastobasids are always scavengers. There are
	sugarcane pests and species intercepted from fresh fruit. The food habits of this
	family require further study.
	There are no keys to blastobasid larvae, but New World interceptions from
	yucca pods can be named if the larvae are large. Sometimes blastobasids from
	acorns, coffee or sugarcane can also be identified. Otherwise leave them at the
	family level. Note in the comments if sclerotized rings are present on A1-8
	(these are also found in Xylorytidae and a few other gelechioids).
•	head with six stemmata, SD1 rings present or absent on A1-8, living and
	dead plants, cosmopolitanBlastobasidae
•	two stemmata present, SD1 rings absent, stored grains, bulbs,
	cosmopolitan Endrosis sarcitrella
48'. Su	ibmentum lacks a large oval pit; A1-8 lacks a sclerotized ring around SD1;
	various hosts, cosmopolitan49
40 T 4	
49. L1	and L2 of A1-8 closely spaced, often on the same pinaculum
	This group includes Gelechioidea, Argyresthiidae, Cossidae, Sesiidae and
	Castniidae. Minet (1991) illustrated certain sensory pits on the head that appear
	unique to cossids and its relatives. In addition, he noted most sesiids, and some
	cossids and castniids, have an oblique line on the prothoracic shield between
	the D setae. Another characteristic of these borers (Cossidae, Sesiidae and
	Castniidae) is that the cuticular microspines are not distributed uniformly
	(Minet 1991), this is unusual. Stehr (1987) noted that SD2 is usually caudal to
	SD1 on the prothorax in Cossidae; this distinguishes cossids from most sesiids.
	Paysandisia archon is the most likely species of castniid to be intercepted from
	Europe but there are many other species of this family in Latin America.
	Chilecomadia are imported into the United States under the name butterworms.
	dorsum with spined areas on A1-8; no crochets; palms, banana, Latin America Castniidae
•	dorsum with oval spined area; prolegs absent but may have spines or a few
	crochets; palms, Europe, Brazil and temperate South America
	one extra setae behind spiracle on A1-8; crochets biordinal; Europe
•	several extra setae mostly above spiracle on A1-8; crochets uniordinal; garlic or
	onions, Italy, the Middle East
•	prothorax smooth; no secondary setae in subdorsal area of A1-8; Canada
	possibly Prionoxystus robinae

• rugose area of prothoracic shield with an anterior triangular patch of	
microspines; cosmopolitan	Zeuzera
 rugose area of prothoracic shield with blunt projections covering almost all of 	
posterior portion; reed mat from Europe	castaneae
• anal shield with pointed horn; bottles of mezcal from Mexico	Comadia
• crochets in transverse band; faint slanted line on prothorax borers in woody	
plants	Sesiidae
• squash, Mexico	Melittia
49'. L1 and L2 of A1-8 widely spaced, not on the same	
pinaculum (Yponomeutoidea & Tineidae)	52
50. Prothoracic shield with SD2 slightly behind a line connecting SD1 and XD2;	
crochets uniordinal; A9 with setae D1, D2, and SD1 on common	
pinaculum; on Sorbus or MalusArgyrest	hia conjugella
Weisman (1986) thought the position of SD2 on the prothoracic shield might	
help define the Argyresthiidae, but according to the drawings in Stehr (1987),	
the position of SD2 varies among species in this family. Argyresthia pruniella	
or Argyresthia eugeniella (West Indies, guava) might key out here too but	
larvae of either were not available.	
50'. Prothoracic shield with SD2 far behind line connecting SD1 and XD2;	
crochets variable; A9 with setae D1, D2, and SD1 not on common	
pinaculum; various hosts	51
51. SV group of A1 bisetose; at least D1 and SD1 in a vertical line and D2 on its	
own pinaculum, either dorsad or posterodorsad of D1; anal comb	. ~
sometimes present mo	st Gelechiidae
Other characters present in some, but not all Gelechiidae, are: mandible with an	
outer (extra) tooth, crochets divided into two groups, and presence of a	
sclerotized collar on the abdominal prolegs. A few species have D1	
anteroventrad of D2 on A9, but this is not common. SD1 on A9 is either	
hairlike or setaform. There is a great need to evaluate many gelechiid	
interceptions and decide which names can be accepted without rearing or	
barcoding as neither of these procedures seem possible in the near future. We	
present a conservative list below. For solanaceous feeding gelechiids, consult	
the dichotomous key in Hayden et al. (2013) for additional species and	
characters. A simplified version of that key is presented below.	
• abdominal prolege rudimentary with only 7 to 4 crochete in stored products	
• abdominal prolegs rudimentary with only 2 to 4 crochets; in stored products	1 11
only; cosmopolitan but rarely intercepted as larvae	cerealella
only; cosmopolitan but rarely intercepted as larvae	cerealella
only; cosmopolitan but rarely intercepted as larvae	cerealella
only; cosmopolitan but rarely intercepted as larvae	cerealella
only; cosmopolitan but rarely intercepted as larvae	
only; cosmopolitan but rarely intercepted as larvae	
 only; cosmopolitan but rarely intercepted as larvae	
 only; cosmopolitan but rarely intercepted as larvae	
 only; cosmopolitan but rarely intercepted as larvae	

setaform, not hairlike; prolegs with greater than ten crochets in a uniordinal penellipse; cotton, okra, and other malvaceous plants, India, Egypt, Latin • head dark brown and typically reticulated, adfrontal setae widely separated. AF2 at level of apex of front; prespiracular group of prothorax not sclerotized; thorax with dark pigmentation; anal comb present; from unknown whole leaf spices, *Gnetium*, *Pterocarpus* and Malvaceae, Africa Dichomerinae • head with adfrontal setae widely separated, AF2 at level of apex of front; prespiracular group of prothorax strongly sclerotized; thorax lacks dark pigmentation;; mature larva with yellow to light brown setae; anal comb present; on an unknown whole leaf spice, Gnetum africanum, Erythrina • head with adfrontal setae close together, AF2 well below apex of front; line joining setae Ll and S2 tangent to or passing through stemma one; thoracic legs pale; lateral setae of A9 in a nearly vertical line; Capsicum and Solanum, New Worldprobably Symmetrischema spp. • head with adfrontal setae close together, AF2 well below apex of front; all L setae of T1 joined on a single sclerotized, sometimes faint, pinaculum; crochets • head with adfrontal setae close together, AF2 well below apex of front; all L setae of T1 on separate pinacula; Solanum, Capsicum, Datura, Nicotiana, • head with adfrontal setae close together, AF2 well below apex of front; L group usually bisetose on A9, occasionally trisetose on one side; mandible triangular with three teeth and the mandibular setal bases in a diagonal line; D1 and D2 pinacula of mesothorax joined; SD1 on A9 • head with adfrontal setae close together, AF2 well below apex of front; mandible with four to five teeth, outer surface of mandible with basal ridge above the condyles (best viewed ventrally), the mandibular setal bases arranged vertically; thoracic legs pigmented; L group trisetose on A9, the L setae in a triangular arrangement; SD1

D2 variable; anal comb absent......MISCELLANEOUS GELECHIOIDEA

on A9 thin and hair-like; in field or stored potatoes,

Separation of all larval Gelechiidae from all Cosmopterigidae is virtually impossible and there is no set of characters to define larval Oecophoridae (in the sense of Weisman 1986). Stathmopodidae and Batrachedridae are two more families that may be common, but are difficult to recognize even to family. Elachistidae is a diverse group that can only be defined in the pupal stage. We can identify some pests but interceptions of Gelechioidea from poorly studied regions like South America, Africa and most of Asia can hardly be placed to family, especially if not associated with economically important plants. They are usually dumped into Oecophoridae for lack of a better placement. Another option is to just leave them at superfamily Gelechioidea. The interceptions from *Brunia* fit this category, they are named either Gelechiidae or Oecophoridae but

no one knows the correct name for sure without rearing or barcoding (assuming there is even a barcode match for our species).

New World *Anatrachyntis* (used to be called *Pyroderces* or *Sathrobrota*) are common but the two species cannot be told apart. Pest lists favor *Anatrachyntis rileyi*, but this may just be an artifact of *A. badia* being poorly known. Some Old World species are still in *Pyroderces*, but there is no morphological informtion on their larvae. *Anatrachyntis* also occur in Europe. Do not confuse *Stenoma catenifer* with the tortricid *Cryptophasma* (also in avocado seeds from Mexico). Many Stenomatinae tie avocado leaves, most of them poorly known.

- Apiaceae, artichoke, pupae (if present) have lateral condyles, Europe and Middle East Elachistidae
- D1 anterodorsad of D2 and SD1 on A9; SD1 of A9 setaform.....poss. Cosmopterigidae

52. Pinaculum of SD1 touching or enclosing spiracle on A1-8; crochets of A3-6 in a uniordinal circle enclosing a short longitudinal series of crochets;

Three species of Acrolepiidae are recorded from *Allium* by Gaedike (1997). They are: *A. assectella*, *A. sapporensis*, and *A. betulella*. The record for *A. assectella* in Hawaii actually refers to *A. sapporensis* (Gaedike 1997). Nevertheless, it is still helpful to consult Zimmerman's summary of the literature. In Europe and Russia, *A.* assectella can be confused with *A. betulella*. Although it seems confusing that *A. betulella* should feed on *Allium ursinum* and not *Betula* (birch), this situation arose because *A. ursinella* was synonymized under *A. betulella* (Gaedike 1997) which is an older more valid name. *Acrolepiopsis sapporensis* occurs from Mongolia to Japan, and was introduced into Hawaii (Gaedike 1997). Specimens of *A. sapporensis* were

and sapporensis was known. If this series included larvae, contact Steven	
Passoa. Thus, interceptions from Canada can be identified as A. assectella and	
those from Hawaii are A. sapporensis. Other orgins can go to genus	
Acrolepiopsis as long as the host is Allium.	
52'. Pinaculum of seta SD1 on A1-8 not touching or enclosing spiracle; crochets in	
circle or ellipse without enclosing series of crochets	53
53. Crochets of abdominal proleg in a biserial circle; seta L3 missing on	
A9PLUTELLIDAE and PRAYDID	AE
There are three pest species of <i>Prays</i> : citri, (citrus, southern Europe),	
endocarpa (citrus, Asia) and oleae (olive leaves or fruit, southern Europe).	
Prays has been in Plutellidae, Yponomeutidae and currently its own family,	
Praydidae.	
• prothoracic and anal shields spotted; body setae dark and thick; AFa absent;	
seven setae on A9 with D, SD and L setae all widely separated, SD1 hairlike;	
anal prolegs longer than broad, with few crochets; Brassicaceae and	
Capparidaceae, cosmopolitan	
• A9 with dorsal and subdorsal setae on one continuous pinaculum, the L setae	
on another pinaculum; SD1 not hairlike; anal prolegs short with many crochets Prays spp.	
53'. Crochets of abdominal prolegs in a uniserial circle or ellipse, rows of very	
small spinules may be present on the prolegs anterior and/or posterior to	
the crochets; seta L3 usually present on abdominal segment 9TINEID	ΑE
We can easily recognize Acrolophinae to subfamily and the genus <i>Opogona</i> . If	
the larvae are from stored products, and slide mounted, the key by Hinton	
(1956) can be tried. The head may need to be cleared and slide mounted to tell	
how many stemmata are present. Identification of tineids associated with green	
plants is difficult because so little is known about the larval morphology of	
anything except the pest species. L3 is missing on A9 in Tineola, but this	
species makes a case and would key out in couplet 12.	
 Head with six stemmata; prespiracular shield surrounds spiracle and is at least 	
partially fused to prothoracic shield; hypostoma (postgena) in broad contact;	
abdominal prolegs with space between the spinules and crochets	
 head with two stemmata; prespiracular shield surrounds spriacle but is not 	
fused to prothoracic shield; abdominal prolegs without space between the	
spinules and crochets	

labelled *A. assectella* and distributed to all the APHIS ports as part of the Pests Not Known to Occur [in the U.S.] (PNKTO) project. Thus, APHIS identifers need to find these specimens (usually marked with a red label) and change the

identification from A. assectella to A. sapporensis. It appears all these examples were collected in Hawaii before the distinction between assectella

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Literature cited in this key is listed at:

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